# Dofetilide Unmasks Occult Congenital Long QT Syndrome Type 2: A Simulation Study

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#### **Abstract**

Accurate diagnosis of long QT syndrome is a key factor for reducing the risk of cardiac arrhythmias. Our goal is to investigate the potential use of dofetilide to unmask latent  $I_{Kr}$  mutation carriers.

A modified version of the O'Hara et al. model was used to simulate the electrical activity of isolated cardiac cells. The original  $I_{Kr}$  formulation was replaced by the Fink et al. Markov model of the human  $I_{Kr}$  channels and our dynamic model of dofetilide was used to simulate drug administration. A sensitivity analysis was performed to study the effect of  $I_{Kr}$  transition rate alterations on AP duration (APD) prolongation in the absence and in the presence of dofetilide.

Our results show that acceleration of the rate transition from open to the last closed state ( $\beta\beta$ ) produced the shortest prolongation of the APD in the absence of the drug. However,  $\beta\beta$  acceleration provoked the highest additional APD prolongation under dofetilide exposure related to the APD prolongation observed before the drug application. In addition, this  $I_{Kr}$  alteration was the transition rate modification that most increased the rate of deactivation.

In conclusion, our observations indicate that dofetilide could potentially be used to unmask  $I_{Kr}$  mutations accelerating the deactivation process.

#### 1. Introduction

Drug-induced or acquired long-QT syndrome (LQTS) is a pathology characterized by prolonged ventricular repolarization subsequent to drug administration [1]. This disorder can lead to potentially mortal arrhythmias, such as torsade de pointes [1].

In the last decades, it has been shown that genetic factors may increase the risk of drug-induced LQTS and arrhythmias [1-3]. Gene mutations have been identified in 10% to 15% of subjects with drug-induced LQTS [3].

Accurate diagnosis of LTQS is essential for reducing the risk of cardiac arrhythmias [4]. Unfortunately, diagnosis based on the QT prolongation is not completely reliable [5] and genetic testing has some disadvantages, such as its difficulty, cost and accessibility [6]. Recently, drug-induced provocative tests have been used to reveal some kinds of latent mutation carriers. In these tests, the QT interval of the subject is measured after the addition of a specific drug. The drug enhances the functional effects of the presumed defective ionic ion channel, which helps to discriminate between normal and mutation carrier subjects.

To investigate the potential use of dofetilide to unmask latent  $I_{Kr}$  mutation carriers a sensitivity analysis of the AP duration (APD) in the absence and in the presence of dofetilide to changes in the  $I_{Kr}$  transition rates was performed. Moreover, the effects of the  $I_{Kr}$  transition rates on the electrophysiological characteristics of  $I_{Kr}$  were investigated.

### 2. Methods

The five-state Markov model defined by Fink et al. [7] was used to simulate the human ventricular  $I_{Kr}$ . This Markov chain is composed of three closed states (C3, C2 and C1), a conducting open state and (O) an inactivation state (I), as depicted in Figure 1A. This  $I_{Kr}$  model was incorporated into the O'Hara et al. human ventricular action potential (AP) model [8] and it was scaled to yield the same  $I_{Kr}$  peak value as the original O'Hara model at 1Hz. Our dynamic model of dofetilide [9] was used to simulate drug administration (Figure 1B).

A sensitivity analysis of the AP duration (APD) in the absence and in the presence of 10 nM dofetilide to changes in the  $I_{Kr}$  transition rates was carried out. It was performed by multiplying or dividing by 5 one  $I_{Kr}$  transition rate at a time. Steady-state APDs at 90 % repolarization were calculated in virtual isolated endocardial cells at 1Hz, although in some cases it was also computed in mid-myocardial cells.

The effects of changes in the  $I_{Kr}$  transition rates on the electrophysiological characteristics of this current were investigated by applying the voltage-clamp protocols defined in [10].

A
$$C_{3} \stackrel{\alpha}{\rightleftharpoons} C_{2} \stackrel{\alpha_{in}}{\rightleftharpoons} C_{1} \stackrel{\alpha\alpha}{\rightleftharpoons} O \stackrel{\alpha_{i}}{\rightleftharpoons} I$$
B
$$C_{3} \stackrel{\alpha}{\rightleftharpoons} C_{2} \stackrel{\alpha_{in}}{\rightleftharpoons} C_{1} \stackrel{\alpha\alpha}{\rightleftharpoons} O \stackrel{\alpha_{i}}{\rightleftharpoons} I$$

$$C_{3} \stackrel{\alpha}{\rightleftharpoons} C_{2} \stackrel{\alpha_{in}}{\rightleftharpoons} C_{1} \stackrel{\alpha\alpha}{\rightleftharpoons} O \stackrel{\alpha_{i}}{\rightleftharpoons} I$$

$$C_{3} \stackrel{\alpha}{\rightleftharpoons} C_{2} \stackrel{\alpha_{in}}{\rightleftharpoons} C_{1} \stackrel{\alpha\alpha}{\rightleftharpoons} O \stackrel{\alpha_{i}}{\rightleftharpoons} I$$

Figure 1. Markov chain of the human  $I_{Kr}$  channels in the absence (A) and in the presence of dofetilide (B).

### 3. Results and discussion

The simulated effects of changes in the I<sub>Kr</sub> transition rates on the simulated human ventricular AP at 1 Hz are shown in Figure 2. This figure depicts the time course of the simulated endocardial AP under control conditions (thick lines) and when each IKr transition rate is fivefolded (thin lines). Our results show that the transition rates between the open and inactivated states ( $\alpha_i$  and  $\beta_i$ ) exert a critical influence on the AP (Figure 2D), followed by the transition rate from the leftmost to the intermediate closed state (\(\beta\_{in}\)) (Figure 2B). As expected, acceleration of the transition rate from the open to the inactivated state  $(\alpha_i)$  and the transition rate from the rightmost to the intermediate closed state  $(\beta_{in})$  prolonged the AP. This effect is due to the fact that these alterations reduce the probability of the conducting open state and, therefore, I<sub>Kr</sub>. Conversely, acceleration of the transition rate from the inactivated to the open state (β<sub>i</sub>) shortened the AP (Figure 2D), as it decreases the probability of the open state. The five-fold increase of the remaining transition rates had a minimal impact on the simulated AP (Figure 2).

Addition of 10 nM of dofetilide, which is in line with its therapeutic dose [11], to WT and mutated cells with one transition rate multiplied or divided by five was simulated to investigate the differential effect of

dofetilide in these cells. Figure 3 illustrates the APD<sub>90</sub> prolongation relative to WT of the simulated mutated cells with longer APD than wild-type (WT) under drugfree conditions (black) and during 10 nM dofetilide exposure (gray). This figure shows that the APD<sub>90</sub> differences between WT and mutated cells observed under drug-free conditions are specially enhanced during 10 nM dofetilide exposure in the cells whose transition rate from the open to the inactivated state  $(\alpha_i)$  is accelerated and those whose transition rate from the inactivated to the open state  $(\beta_i)$  is slowed. This effect is also observed in those cells whose transition rate from the open to the last closed state (ββ) is accelerated or whose transition rate from the last closed state to the open state  $(\alpha\alpha)$  or the transition rate from the intermediate to the rightmost closed state (ain) is decreased. Comparison of the additional increments of the APD observed in the mutated cells in the presence and in the absence of dofetilide reveals that this drug preferentially

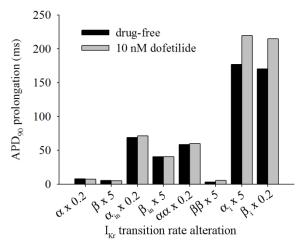


Figure 3. Influence of  $I_{Kr}$  transition rates on the endocardial APD under drug-free conditions (black) and during 10 nM dofetilide exposure (gray). The y-axis represents the APD $_{90}$  prolongation relative to WT. Only those  $I_{Kr}$  transition rate alterations that prolonged the APD are represented.

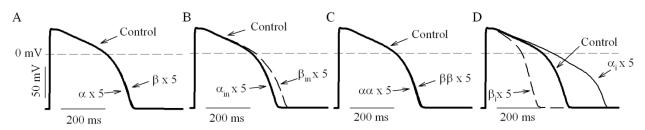


Figure 2. Time course of the simulated endocardial AP under control conditions (thick lines) and when each  $I_{Kr}$  transition rate is five-folded (thin lines), namely,  $\alpha$  (solid line) and  $\beta$  (dashed line) (A);  $\alpha_{in}$  (solid line) and  $\beta_{in}$  (dashed line) (B);  $\alpha\alpha$  (solid line) and  $\beta\beta$  (dashed line) (C);  $\alpha_i$  (solid line) and  $\beta_i$  (dashed line) (D).

enhances the functional effects of the acceleration of the transition rate from the open to the rightmost closed state (ββ). Specifically, the APD difference between endocardial WT cells and those affected by the acceleration of the transition rate from the open to the rightmost closed state (ββ) (3ms) was doubled (6 ms) when 10 nM dofetilide was added (Figure 3 and Figures 4A and 4B). When 30 nM dofetilide was applied it increased five-fold (15 ms). As the effect of this rate on the APD under drug-free conditions is very small, our results suggest that dofetilide could be used to unmask this kind of genetic defect. As mid-myocardial cells are more sensitive to defective I<sub>Kr</sub>, the effect of the five-fold increase of the transition rate from the open to the rightmost closed state (ββ) was also investigated in this type of cells. Our simulations show that the APD difference between mid-myocardial WT cells and those affected by the acceleration of the transition rate from the open to the rightmost closed state (ββ) (4 ms) was increased seven-fold (28 ms) under 10 nM dofetilide (Figures 4C and 4D).

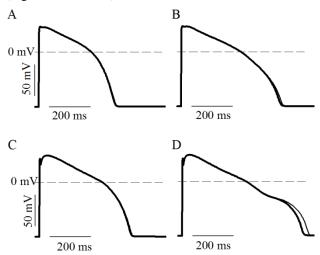


Figure 4. Steady-state AP time-course of WT (thick lines) and mutated cells with the transition rate from the open to the rightmost closed state rate five-folded (thin lines) in isolated endocardial (top row) and mid-myocardial (bottom row) cells in the absence (left column) and in the presence of 10 nM dofetilide (right column).

Figure 5 shows the characterization of WT and mutant  $I_{Kr}$  channels with a five-fold increase of the transition rate from the open to the rightmost closed state ( $\beta\beta$ ). This rate alteration significantly reduced the deactivation time constant (Figure 5C). Specifically, this time constant was approximately divided by five at very negative membrane potentials. It also caused an 8.7 mV leftward shift of the steady-state activation curve (Figure 5A) as well as a 7.4 mV rightward shift and a 10 % decrease of the slope of the steady-state inactivation curve (Figure 5B). A sensitivity study of the electrophysiological

characteristics of IKr to changes in its transition rates revealed that the transition rate from the open to the rightmost closed state ( $\beta\beta$ ) was the most relevant for the deactivation time constant, followed by the transition rates between the open and inactivated state ( $\alpha_i$  and  $\beta_i$ ), although their influence was much more smaller. In addition, the impact of the transition rate from the open to the rightmost closed state  $(\beta\beta)$  on the steady-state activation and inactivation curves was significantly smaller than the influence of other transition rates. Therefore, our observations indicate that dofetilide could potentially be used to unmask I<sub>Kr</sub> mutations accelerating the deactivation process. This observation is in accordance to the clinical and experimental reports of patients with normal QT interval, but affected by mutations accelerating the deactivation of IKr, that suffered drug-induced arrhythmia [1,2,12,13].

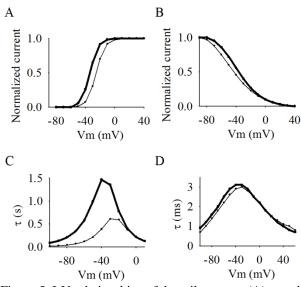


Figure 5. I-V relationships of the tail currents (A), steady-state inactivation curves (B), activation and deactivation time constants (C) and inactivation and recovery from inactivation time constants for WT (thick lines) and mutant (thin lines)  $I_{Kr}$  channels with a five-fold increase of the transition rate from the open to the rightmost closed state ( $\beta\beta$ ).

## 4. Conclusions

Our simulations predict that the transition rates between the open and inactivated state of  $I_{Kr}$  are pivotal for APD at 1Hz. In addition, the transition rate from the open to the rightmost closed state has a minimal effect on APD. However, under dofetilide exposure de influence of this rate on the APD is specially amplified. Moreover, it was the transition rate modification that most accelerates the deactivation process. Therefore, our results suggest that dofetilide could potentially be used to unmask  $I_{Kr}$ 

mutations accelerating the deactivation process.

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